

HPV INFECTION AND MALIGNANCY: CURRENT UNDERSTANDING AND FUTURE PERSPECTIVES

Infecção por HPV e malignidade: conhecimento atual e perspectivas futuras

Jair C. Leão¹
Giusepina Campisi²
Jose Ricardo D. Pereira¹
Alessandra A. Tavares Carvalho¹

Abstract

Human papillomaviruses (HPV) are a group of genetically related organisms that commonly infect stratified squamous epithelium. Unlike many other viruses that infect epithelium and induce lysis of the cells they induce proliferative changes that result in both benign and malignant tumors. The aim of the present paper was to briefly review current understanding of HPV infection in relation to malignancy and also highlight perspectives with regard to the development of an effective vaccine.

Keywords: Human papillomavirus; Oral pathology; Malignancies.

Resumo

O papilomavírus humano (PVH) constitui um grupo de organismos geneticamente relacionados que freqüentemente infectam o epitélio escamoso estratificado. Ao contrário de muitos outros vírus que infectam o epitélio e induzem lise das células, eles induzem alterações proliferativas que resultam em tumores malignos e benignos. O objetivo do presente artigo foi revisar brevemente o atual conhecimento da infecção por HPV em relação com malignidade e também ressaltar as perspectivas com relação ao desenvolvimento de uma vacina efetiva.

Palavras chave: Papilomavírus humano; Patologia bucal; Neoplasias malignas.

¹ Departamento de Clínica e Odontologia Preventiva, Universidade Federal de Pernambuco, Recife, Brazil. Address Dr. Jair C. Leao, Departamento de Clínica e Odontologia Preventiva Universidade Federal de Pernambuco, Av. Prof. Moraes Rego, 1235. Recife, PE, Brazil. CEP 50670-901. jleao@ufpe.br

² Dip. Scienze Stomatologiche, University of Palermo, Palermo, Italy.

Rev. de Clín. Pesq. Odontol., v. 1, n.1, jul./ago. 2005

Introduction

To date, more than 100 types of human papillomavirus (HPV) have been identified. In the past 20 years, there has been an increasing interest in HPVs due to their potential role in the pathogenesis of malignant tumors. HPV infections are known to affect predominantly adult, sexually active age groups, whereas skin warts, at various anatomic sites, are usually associated with younger individuals (1).

In fact, HPV are long thought to be a common sexually transmitted viral infection. The virus targets injured cervical epithelial cells, gaining access at sites of microtrauma occurring during (sexual) intercourse. In addition, a subset, termed high risk types, may be strongly associated with the development of oral and cervical cancer (2,3). HPV16 is the most common HPV type found in ano-cervical cancers, where it is detected in over 50% of cases, while other HPV types commonly observed in cervical cancers include types 18, 45 and 31.(4) Similarly, HPV16 is the most commonly detected type in oral and pharyngeal carcinomas(5). In the mouth, HPV has been associated with cancer despite other risk activities, such the use of tobacco and/or alcohol (6). One possibility is that P53 (gene) inactivation is a major component of HPV-related carcinogenesis (7).

Furthermore, it has been shown that a single patient may concomitantly suffer from both cervical and oral cancer, although molecular biology techniques may prove useful to differentiate primary and secondary lesions. In one study, for instance, a patient with apparent mouth metastasis from cervical cancer was found to harbor two HPV sequences from different viruses (18 and 33, for the uterine cervix and the mouth, respectively), thus indicating the oral lesion as a synchronous second primary tumor.(8)

Viral structure

HPV's are small, nonenveloped DNA viruses that replicate in the nuclei of squamous epithelium cells. The viral genome is encased in a capsid layer, consisting of a major structural protein and a minor structural protein.(9) All of the potential coding regions, or open reading frames

(ORFs), exist in one of the two DNA strands (10). This means that all of the genetic information is located in only one strand.

The genome is divided in three regions; a long coding region, and two regions consisting of the designated ORFs, called the early and late regions (11) The long coding region contains the viral origin for replication. It is responsible for the regulation of replication, and controls the transcription of some gene sequences in the early region. The early region, divided in eight coding regions, encodes important proteins in viral replication occurring 'early' in the viral life cycle. It is also responsible for maintaining high number of HPV and for high-risk types immortalization. The late region encodes for viral structural proteins necessary for capsid production (12).

HPV life cycle

HPV infection primarily infects the basal layer of the injured epithelium. The viral genomes are established in the host cell as unintegrated extrachromosomal elements or episomes. After infection, the first viral genes to be expressed are E1 and E2, which are the replication proteins. These proteins bind to the origin of DNA replication, located on sites within the long coding region (13). In the basal layer, the virus is in a nonproductive stage and is present in low copy numbers. The virus proliferates here, recruiting host factors for viral synthesis, and replicating its DNA enough to keep up with the mitosis of basal cells (9).

As the host cells continue their normal life cycle or pattern of maturation, a subset of daughter cells detach and migrate from the basal layer. The infected host cells divide, and HPV DNA is divided between the daughter cells as they stratify and differentiate. Virus DNA travels with the host cells as they undergo their normal life cycle and mature. HPV does not encode a DNA polymerase and, hence, it is dependent on host cell differentiation to continue its own life cycle. The virions proliferate, moving with the host cells toward the terminally differentiating or keratinizing layers of the epithelium (9).

Differentiation

HPV-infected cells mature, stratify, and develop special characteristics during a process called differentiation. E4 is a protein expressed later in these terminally differentiating cells, and is also found in association with the viral capsids. Late region genes are also expressed in the differentiated cells near the surface of the epithelium, initiating the synthesis of the capsid layer for the genome. The surface epithelial cells do not divide, but their location offers an ideal site for viral transmission. The shedding of the naturally dying surface cells laden with HPV, also called koilocytes, serves as the vector of transmission. Once the capsid protein layer is formed, the genome is capable of infection and transformation(14).

Viral Integration

Incorporation of HPV genomes in the human host DNA, called integration, appears to be one of the final steps toward malignant transformation. This occurs in the HPV-16, and other high-risk types, but in low-risk types the viral genome remains as extrachromosomal plasmids.

Integration in the host cell DNA occurs in a break in the viral genome, usually at E1 and E2 region. The loss of host cellular control, and the persistence of the high risk virion may contribute to this occurrence. The disruption at E1/E2 plays a pivotal role in oncogenesis.(15) Integration at this region leads to the activation of p97, the normally suppressed papillomaviral promoter that directly expresses E6 and E7. These two viral transforming genes possess oncogenic properties; they have the capacity to functionally inactivate the cellular tumor suppressors p53, and retinoblastoma protein (pRb). E6 complexes with p53, and E7 with pRb, thereby interrupting the existing cellular pathways that would normally lead to growth arrest and cell death. Integration causes the increased expression of E6 and E7, providing a selective growth advantage to the affected epithelial cell, but it alone is not sufficient for malignant transformation.(16)

HPV detection

Human papillomavirus is an ubiquitous or commensal entity responsible for skin infection that have a worldwide distribution

with a broad spectrum of genotypes. The prevalence of HPV DNA varied from 42% in Zambia to 70% in Sweden (17). Two or multiple genotypes were frequently found in the same sample and the most prevalent HPV type is thought to be HPV-5, with an overall prevalence of 6.5 %. This was also the only type that was found in samples from all of the countries in that study.(17)

The prevalence of HPV DNA among children from the ages of 1 month to 4 years varies between 50 and 70%. The HPV DNA sequences commonly detected suggest a great diversity of genotypes and putative genotypes. In a study, (18) among 115 samples, a total of 73 HPV types or putative types were isolated. Of these, 26 putative HPV types had not been described before. Hence, asymptomatic HPV infections of normal skin seem to be acquired very early in infancy and are caused by a great multiplicity of HPV types.

HPV-DNA positivity in the absence of clinically or colposcopically detected lesions is a rare event. It has been shown that HPV DNA was detected in only one sample (3%) from men without visible lesions, in 5 samples (15%) from men with penile lesions but without urethral lesions, and in 16 men with urethral lesions (78%).(19) On the other hand, men aged 18-70 years attending a sexually transmitted disease clinic were screened for the presence of HPV infection. Penile skin swabs were assessed for HPV DNA using polymerase chain reaction with reverse line-blot genotyping and the prevalence of HPV was 28.2%. Oncogenic HPV types in that study were found in 12.0% of participants, nononcogenic types were found in 14.8% of participants, multiple types were found in 6.1% of participants, and unknown types were found in 5.9% of participants. The most prevalent subtypes were the nononcogenic varieties 6, 53, and 84. HPV positivity was not associated with age. These results indicate that HPV infection among men at high risk is common but that characteristics of male HPV infection may differ from those of female infection.(20) HPV infection can be detected not only by DNA amplification, but also using serological methods. However, the poor sensitivity of anti-HPV IgG would suggest the need of new methods of HPV detection: in fact, in a recent research, only

50% of HPV DNA positive individuals were also HPV-seropositive (21).

The lack of sensitivity may also explain the reason for the low detection rate of HPV antibodies in a large retrospective serological cohort, where only 47% of sera from women with cervical cancer and 33% of individuals with oropharyngeal disease were HPV16 L1 positive.(22)

Nevertheless, it has been estimated that HPV seroprevalence in Thailand women varies from 3.9 to 9.1%, with the highest prevalence rates associated with younger age, herpes simplex type 2 (HSV-2) infection and for females being married with partners who have extra-marital sexual partners.(23)

HPV and other malignancies

HPV has been more controversially associated with prostate cancer;(24) squamous cell and adenosquamous carcinoma of the colon and rectum;(25;26) ovarian carcinoma;(27) squamous cell carcinoma of the fingers;(28;29) non-melanoma skin malignancies such as basal cell carcinoma;(30) anal cancer and its precursor lesion, anal squamous intraepithelial lesions(31) and transitional cell carcinoma (TCC) of the urinary bladder (32;33), although these results are not supported by others who have not found association of HPV types 6, 11, 16, 18, 31, 33 and 51 with carcinoma of the bladder.(34) In addition, HPV DNA has been detected in a small proportion of cases of bronchopulmonary carcinoma, and thus HPV infection appears to play a limited role in the tumorigenesis of most lung carcinomas.(35).

There is an increasing evidence supporting the role of HPV in the development of squamous cell carcinoma of the head and neck (SCCHN). Recently, some large sample (36-41) size studies showed that SCCHN and potentially malignant oral lesions, histologically diagnosed, contain DNA of high-risk HPV genotypes (e.g. HPV-18 and -16).

HPV and cancer prognosis

HPV has been found to be an indicator of the severity of oral cancer.(42) In fact, it had

already been observed that HPV DNA was associated with all confirmed grade 3 cervical intraepithelial neoplasia and primary cervical cancers.(43) It may be that the histological grading of cervical lesions are not only associated with HPV type, but also to HPV DNA viral load.(44)

In addition, it is possible that polymorphisms in human leukocyte antigen (HLA) genes are implicated in the risk for developing human papillomavirus (HPV)-associated cervical neoplasia.(45) Also, HPV DNA loads of six oncogenic HPV types were measured in cervical scrapes of human immunodeficiency virus (HIV)-infected and uninfected women. In both groups, HPV loads increased with the grade of cervical disease. In the same study, HIV infection did not affect HPV loads in low-grade lesions but was associated with significantly higher HPV loads in severe dysplasia; highest loads were found in advanced HIV disease.(46)

HPV vaccine

HPV infection may be prevented by neutralizing antibodies specific for the viral capsid proteins. In clinical trials, vaccines comprised of HPV virus-like particles (VLPs) have shown great promise as prophylactic HPV vaccines.(47) However, due to the fact that capsid proteins are not expressed at detectable levels by infected basal keratinocytes, vaccines with therapeutic potential must target other non-structural viral antigens. Two HPV oncogenic proteins, E6 and E7, are important in the induction and maintenance of cellular transformation and are co-expressed in the majority of HPV-containing carcinomas. Therefore, therapeutic vaccines targeting these proteins may have potential to control HPV-associated malignancies.(48) Various candidate to therapeutic HPV vaccines are currently being tested whereby E6 and/or E7 is administered in live vectors, in peptides or protein, in nucleic acid form, as components of chimeric VLPs, or in cell-based vaccines.(49;50) Encouraging results from experimental vaccination studies in animal models have led to several prophylactic and therapeutic vaccine clinical trials.(51-53) Should they fulfill their promise, these vaccines may prevent HPV infection or control its

potentially life-threatening consequences in humans.

References

1. Syrjanen S, Puranen M. Human papillomavirus infections in children: the potential role of maternal transmission. *Crit Rev Oral Biol Med* 2000; 11:259-274.
2. Walboomers JM, Jacobs MV, Manos MM, Bosch FX, Kummer JA, Shah KV et al. Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. *J Pathol* 1999; 189:12- 19.
3. Sugiyama M, Bhawal UK, Dohmen T, Ono S, Miyauchi M, Ishikawa T. Detection of human papillomavirus-16 and HPV-18 DNA in normal, dysplastic, and malignant oral epithelium. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2003; 95:594-600.
4. Bosch FX, Manos MM, Munoz N, Sherman M, Jansen AM, Peto J et al. Prevalence of human papillomavirus in cervical cancer: a worldwide perspective. International biological study on cervical cancer (IBSCC) Study Group. *J Natl Cancer Inst* 1995; 87:796-802.
5. Kreimer AR, Alberg AJ, Daniel R, Gravitt PE, Viscidi R, Garrett ES et al. Oral human papillomavirus infection in adults is associated with sexual behavior and HIV serostatus. *J Infect Dis* 2004; 189:686-698.
6. Smith EM, Ritchie JM, Summersgill KF, Hoffman HT, Wang DH, Haugen TH et al. Human papillomavirus in oral exfoliated cells and risk of head and neck cancer. *J Natl Cancer Inst* 2004; 96:449-455.
7. Dai M, Clifford GM, le Calvez F, Castellsague X, Snijders PJ, Pawlita M et al. Human papillomavirus type 16 and TP53 mutation in oral cancer: matched analysis of the IARC multicenter study. *Cancer Res* 2004; 64:468- 471.
8. Fracchioli S, Porpiglia M, Arisio R, Voglino G, Katsaros D. Oral squamous carcinoma in a patient with cervix cancer: use of human papillomavirus analysis to differentiate synchronous versus metastatic tumor. *Gynecol Oncol* 2003; 89:522-525.
9. Campo MS. Animal models of papillomavirus pathogenesis. *Virus Res* 2002; 89:249-261.
10. ernard HU. Gene expression of genital human papillomaviruses and considerations on potential antiviral approaches. *Antivir Ther* 2002; 7:219-237.
11. eh WL, Middleton K, Christensen N, Nicholls P, Egawa K, Sotlar K et al. Life cycle heterogeneity in animal models of human papillomavirus-associated disease. *J Virol* 2002; 76:10401-10416.
12. obery TW, Caulfield MJ. Identification of Tcell epitopes using ELISpot and peptide pool arrays. *Methods Mol Med* 2004; 94:121-32.
13. ollmann R, Mehes G, Torka R, Speich N, Schmitt C, Bollmann M. Human papillomavirus typing and DNA ploidy determination of squamous intraepithelial lesions in liquid-based cytologic samples. *Cancer* 2003; 99:57-62.
14. eley S. Human papillomavirus: beware the infection you can't see. *Aust Fam Physician* 2003; 32:311-315.
15. orng PL, Chan WY, Lin CT, Huang SC. Decreased expression of human papillomavirus E2 protein and transforming growth factorbeta1 in human cervical neoplasia as an early marker in carcinogenesis. *J Surg Oncol* 2003; 84:17-23.
16. inawaer, Ahmatjan A, Suzuk L. [Detection of HPV type 16, 18 infection and p53 protein overexpression in oral squamous cell carcinoma]. *Zhonghua*

- Kou Qiang Yi Xue Za Zhi 2001; 36:451-453.
17. ntonsson A, Erfurt C, Hazard K, Holmgren V, Simon M, Kataoka A et al. Prevalence and type spectrum of human papillomaviruses in healthy skin samples collected in three continents. *J Gen Virol* 2003; 84:1881-1886.
 18. ntonsson A, Karanfilovska S, Lindqvist PG, Hansson BG. General acquisition of human papillomavirus infections of skin occurs in early infancy. *J Clin Microbiol* 2003; 41:2509- 2514.
 19. ynaud O, Ionesco M, Barrasso R. Cytologic detection of human papillomavirus DNA in normal male urethral samples. *Urology* 2003; 61:1098-1101.
 20. aldwin SB, Wallace DR, Papenfuss MR, Abrahamsen M, Vaught LC, Kornegay JR et al. Human papillomavirus infection in men attending a sexually transmitted disease clinic. *J Infect Dis* 2003; 187:1064-1070.
 21. iscidi RP, Ahdieh-Grant L, Clayman B, Fox K, Massad LS, Cu-Uvin S et al. Serum immunoglobulin G response to human papillomavirus type 16 virus-like particles in human immunodeficiency virus (HIV)-positive and risk-matched HIV-negative women. *J Infect Dis* 2003; 187:194-205.
 22. an Doornum GJ, Korse CM, Buning-Kager JC, Bonfrer JM, Horenblas S, Taal BG et al. Reactivity to human papillomavirus type 16 L1 virus-like particles in sera from patients with genital cancer and patients with carcinomas at five different extragenital sites. *Br J Cancer* 2003; 88:1095-1100.
 23. ukvirach S, Smith JS, Tunsakul S, Munoz N, Kesarat V, Opatatian O et al. Populationbased human papillomavirus prevalence in Lampang and Songkla, Thailand. *J Infect Dis* 2003; 187:1246-1256.
 24. dami HO, Kuper H, Andersson SO, Bergstrom R, Dillner J. Prostate cancer risk and serologic evidence of human papilloma virus infection: a population-based case-control study. *Cancer Epidemiol Biomarkers Prev* 2003; 12:872-875.
 25. udeau A, Han HW, Johnston MJ, Whitehead MW, Frizelle FA. Does human papilloma virus have a role in squamous cell carcinoma of the colon and upper rectum? *Eur J Surg Oncol* 2002; 28:657-660.
 26. ee YM, Leu SY, Chiang H, Fung CP, Liu WT. Human papillomavirus type 18 in colorectal cancer. *J Microbiol Immunol Infect* 2001; 34:87- 91.
 27. u QJ, Guo M, Lu ZM, Li T, Qiao HZ, Ke Y. Detection of human papillomavirus-16 in ovarian malignancy. *Br J Cancer* 2003; 89:672- 675.
 28. lam M, Caldwell JB, Eliezri YD. Human papillomavirus-associated digital squamous cell carcinoma: literature review and report of 21 new cases. *J Am Acad Dermatol* 2003; 48:385- 393.
 29. ragg JW, Ratner D. Human papillomavirus type 2 in a squamous cell carcinoma of the finger. *Dermatol Surg* 2003; 29:766-768.
 30. eyer T, Arndt R, Christophers E, Nindl I, Stockfleth E. Importance of human papillomaviruses for the development of skin cancer. *Cancer Detect Prev* 2001; 25:533-547.
 31. iketty C, Darragh TM, Da Costa M, Bruneval P, Heard I, Kazatchkine MD et al. High prevalence of anal human papillomavirus infection and anal cancer precursors among HIV-infected persons in the absence of anal intercourse. *Ann Intern Med* 2003; 138:453- 459.

32. rank GA, Zavalishina LE, Andreeva I. [Immunohistochemical characteristics and a degree of differentiation of urinary bladder cancer]. *Arkh Patol* 2002; 64:16-18.
33. Lopez-Beltran A, Escudero AL, Vicioso L, Munoz E, Carrasco JC. Human papillomavirus DNA as a factor determining the survival of bladder cancer patients. *Br J Cancer* 1996; 73:124-127.
34. Estenend PJ, Stoop JA, Hendriks JG. Human papillomaviruses 6/11, 16/18 and 31/33/51 are not associated with squamous cell carcinoma of the urinary bladder. *BJU Int* 2001; 88:198-201.
35. Iavel CE, Nawrocki B, Bosseaux B, Poitevin G, Putaud IC, Mangeonjean CC et al. Detection of human papillomavirus DNA in bronchopulmonary carcinomas by hybrid capture II: a study of 185 tumors. *Cancer* 2000; 88:1347-1352.
36. Iouda M, Gorgoulis VG, Kastrinakis NG, Giannoudis A, Tsoi E, Danassi-Afentaki D et al. "High risk" HPV types are frequently detected in potentially malignant and malignant oral lesions, but not in normal oral mucosa. *Mod Pathol* 2000; 13:644-653.
37. Gillison ML, Koch WM, Capone RB, Spafford M, Westra WH, Wu L et al. Evidence for a causal association between human papillomavirus and a subset of head and neck cancers. *J Natl Cancer Inst* 2000; 92:709-720.
38. Iovannelli L, Campisi G, Lama A, Giambalvo O, Osborn J, Margiotta V et al. Human papillomavirus DNA in oral mucosal lesions. *J Infect Dis* 2002; 185:833-836.
39. Iingstrom E, Peters E, Hasegawa M, Posner M, Liu M, Kelsey KT. Human papillomavirus type 16 and squamous cell carcinoma of the head and neck. *Clin Cancer Res* 2002; 8:3187-3192.
40. Itchie JM, Smith EM, Summersgill KF, Hoffman HT, Wang D, Klussmann JP et al. Human papillomavirus infection as a prognostic factor in carcinomas of the oral cavity and oropharynx. *Int J Cancer* 2003; 104:336-344.
41. Cully C. Oral squamous cell carcinoma; from an hypothesis about a virus, to concern about possible sexual transmission. *Oral Oncol* 2002; 38:227-234.
42. Smith EM, Ritchie JM, Summersgill KF, Klussmann JP, Lee JH, Wang D et al. Age, sexual behavior and human papillomavirus infection in oral cavity and oropharyngeal cancers. *Int J Cancer* 2004; 92:766-772.
43. Schmeier G, Van den Brule AJ, Brummer O, Meijer CL, Petry KU. No confirmed case of human papillomavirus DNA-negative cervical intraepithelial neoplasia grade 3 or invasive primary cancer of the uterine cervix among 511 patients. *Am J Obstet Gynecol* 2003; 189:118-120.
44. Bba MC, Mouron SA, Gomez MA, Dulout FN, Golijow CD. Association of human papillomavirus viral load with HPV16 and high-grade intraepithelial lesion. *Int J Gynecol Cancer* 2003; 13:154-158.
45. Davidson EJ, Davidson JA, Sterling JC, Baldwin PJ, Kitchener HC, Stern PL. Association between human leukocyte antigen polymorphism and human papillomavirus 16-positive vulval intraepithelial neoplasia in British women. *Cancer Res* 2003; 63:400-403.
46. Eissenborn SJ, Funke AM, Hellmich M, Mallmann P, Fuchs PG, Pfister HJ et al. Oncogenic human papillomavirus DNA loads in human immunodeficiency virus-positive women with high-grade cervical lesions are strongly elevated. *J Clin Microbiol* 2003; 41:2763-2767.

47. andic A, Vujkov T. Human papillomavirus vaccine as a new way of preventing cervical cancer: a dream or the future? *Ann Oncol* 2004; 15:197-200.
48. elters MJ, de Jong A, van den Eeden SJ, van der Hulst JM, Kwappenberg KM, Hassane S et al. Frequent display of human papillomavirus type 16 E6-specific memory t-Helper cells in the healthy population as witness of previous viral encounter. *Cancer Res* 2003; 63:636-641.
49. auser H, Shen L, Gu QL, Krueger S, Chen SY. Secretory heat-shock protein as a dendritic cell-targeting molecule: a new strategy to enhance the potency of genetic vaccines. *Gene Ther* 2004; .
50. allez S, Simon P, Maudoux F, Doyen J, Noel JC, Beliard A et al. Phase I/II trial of immunogenicity of a human papillomavirus (HPV) type 16 E7 protein-based vaccine in women with oncogenic HPV-positive cervical intraepithelial neoplasia. *Cancer Immunol Immunother* 2004; .
51. arcia-Carranca A. Vaccines against human papillomavirus and perspectives for the prevention and control of cervical cancer. *Salud Publica Mex* 2003; 45 Suppl 3:S437-S442.
52. im JW, Hung CF, Juang J, He L, Kim TW, Armstrong DK et al. Comparison of HPV DNA vaccines employing intracellular targeting strategies. *Gene Ther* 2004; .
53. astrana DV, Buck CB, Pang YY, Thompson CD, Castle PE, FitzGerald PC et al. Reactivity of human sera in a sensitive, high-throughput pseudovirus-based papillomavirus neutralization assay for HPV16 and HPV18. *Virology* 2004; 321:205-216.

Received in 07/05/2004; Accepted in 08/12/2004.

Recebido em 05/07/2004; Aceito em 12/08/2004.