Fisiograft™ as a bone graft material in the treatment
of periodontal vertical defects and its clinical
and radiological evaluation: clinical study

Fisiograft® como material de enxerto ósseo no
tratamento de defeitos periodontais verticais e sua
avaliação clínica e radiográfica: um estudo clínico

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Abstract

Objectives: The purpose of this clinical study is to determine the efficacy of Fisiograft™ as a bone graft material in the treatment of three wall vertical defects in generalized chronic periodontitis patients and their clinical and radiological evaluation. Materials and methods: Twenty patients (with 30 defects) diagnosed with generalized chronic periodontitis having two or more three wall vertical defects were selected for this study. Clinical parameters like plaque index, gingival index, probing pocket depth and clinical attachment levels were recorded at different points of time over six months. Radiographic evaluation included the depth of the bone defect and the percentage of bone defect fill, and was carried out for both the groups at baseline, three months and six months. After recording clinical parameters and administering phase-1 therapy, the sites were randomly treated either with Fisiograft™ or open flap debridement only. Results: At the end of six months there was a significant reduction in the plaque and gingival scores in both test and control groups. There was 64% decrease in probing pocket depth for the test site as compared to 55% decrease seen for the control group. Similarly there was an 85% gain in clinical attachment level from the baseline to six months post operatively for the experimental group in comparison to 69% gain for the control group. Furthermore, 44% bone fill was observed for the experimental site whereas only 18% of bone fill was evident in the control site. Conclusion: Fisiograft™ improves healing outcomes, leads to a reduction of probing depth, a resolution of osseous defects and a gain in clinical attachment, compared with open flap debridement by itself.

Keywords: Open flap debridement. Periodontitis. Vertical defects. Polylactic acid. Polyglycolic acid.
Resumo

Objetivo: O objetivo do presente estudo clínico foi determinar a eficácia do Fisiograft®, como material de enxerto ósseo, no tratamento de defeitos ósseos verticais de três paredes em pacientes com periodontite crônica, bem como avaliações clínica e radiográfica. Materiais e métodos: Vinte pacientes (com 30 defeitos) diagnosticados com periodontite crônica generalizada, portando dois ou mais defeitos ósseos verticais de três paredes foram selecionados para o estudo. Parâmetros clínicos como índice de placa, índice gengival, profundidade de bolsa à sondagem e níveis clínicos de inserção foram registrados em diferentes intervalos de tempo até seis meses. Avaliações radiográficas incluíram a profundidade do defeito ósseo e a porcentagem de preenchimento do defeito ósseo, sendo realizadas em ambos os grupos imediatamente (baseline), em três meses e seis meses. Após registrar os parâmetros clínicos e administrar a terapia de fase-I, os locais foram tratados aleatoriamente com Fisiograft® ou retalho de espessura total somente. Resultados: Ao fim do período de seis meses houve redução significativa nos índices de placa e gengival em ambos os grupos, controle e experimental. Houve redução de 64% na profundidade de bolsa à sondagem para os locais de teste comparado, 55% de redução no grupo controle. Similarmente, houve ganho de 85% no nível clínico de inserção do baseline para o período de seis meses de pós-operatório para o grupo experimental em comparação ao ganho de 69% para o grupo controle. Adicionalmente, um preenchimento ósseo de 44% foi observado para os locais experimentais, enquanto somente 18% de preenchimento foi evidente nos locais de controle. Conclusão: O Fisiograft® melhora os resultados de cicatrização, promove redução na profundidade de sondagem, constitui uma solução para os defeitos ósseos e aumenta na inserção clínica, comparado ao retalho de espessura total somente.


Introduction

Periodontal disease is one of the most prevalent diseases worldwide. Bacterial plaque has been implicated as the major etiologic agent in the initiation and progression of inflammatory periodontal disease. Periodontal disease results in a loss of periodontal attachment apparatus, including loss of tooth-supporting alveolar bone. Although periodontitis is an infectious disease of the gingival tissue, changes that occur in the bone are crucial because the destruction of the bone is responsible for tooth loss (1).

The purpose of periodontal therapy is to eliminate the inflammation of the periodontal tissues, to arrest the destruction of soft tissue and bone caused by periodontal disease, and regenerate the lost tissue, if possible (2).

Non-surgical and surgical procedures have been used in periodontal therapy in order to establish healthy periodontal conditions. The non-surgical procedures consist mainly of scaling and root planning and use of chemotherapeutic agents as adjunct, whereas surgical procedures provides access for better and effective instrumentation as well as chance for root biomodification, placement of bone graft or guided tissue regeneration membrane or both.

Non-surgical therapy in deep pockets and furcation areas are not effective in reducing pocket depth and clinical attachment gain than open flap debridement, whereas bone grafting is the most common form of regenerative therapy and has been used for almost 100 years in attempts to stimulate healing of bony defects. The first recorded attempt to use a bone graft was by a Dutch surgeon, Job van Meckren, in 1668. However, the first use of a bone graft to rebuild bone loss by periodontal disease was reported by Hegeduus, in 1923. Materials such as plaster of paris, heterogenous bone powder, and other bone preparations were also tried for implantation into intrabony periodontal defects during the 1930s (3).

A wide range of bone grafting materials, including bone grafts and bone graft substitutes have been applied and evaluated clinically, including autograft, allograft, xenograft and alloplasts (synthetic and semi synthetic materials). Alloplastic bone graft materials are synthetic, inorganic, biocompatible, and bioactive bone substitutes that are believed to promote healing of bone defects through osteoconduction.
Several calcium phosphate biomaterials have been tested since the mid-1970. They have excellent tissue compatibility and do not elicit any inflammation or foreign body response (4, 5).

Polyactic acid (PLA)/polyglycolic acid (PGA) and their copolymers have been successfully used to fabricate bone graft substitutes. As the polymer degrades in vivo, it serves not only as a vehicle for drug delivery, but also as a scaffold to support new bone formation (6). Biodegradable polymers, especially those belonging to the family of polyactic acid (PLA) and polyglycolic acid (PGA), have played an increasingly important role in bone re-constructive procedures. Although extensively used in orthopedics for over a decade, PLA/PGA biomaterials have been scarcely used in cranio maxillofacial applications. In Dentistry, surgical sutures and absorbable membranes in PGA and/or PLA acids have been available for some time for use in guided tissue regeneration. However, only in recent years absorbable synthetic biopolymers have been used as bone fillers in periodontics, proving effective stimulants to bone regeneration. Implantation of PLA-derived devices has been studied to prevent alveolar osteitis or dry socket in extraction sites (7, 8) as well as in the treatment of periodontal osseous defects with access flap alone, PLA implant and decalcified freeze-dried bone allograft (9).

Recently, a new copolymer (Fisiograft™), Ghimas SpA, Casalecchio di Reno, Italy) of 50% DL lactic acid and 50% glycolic acid (50 PLA: 50 PGA) mixed with dextran 125 asexcipient, has been marketed in different formulations, such as sponge, gel, and powder. Being synthetic, it is absolutely free from risk of cross contamination due to pathological agents such as bovine spongiform encephalopathy (BSE), hepatitis and human immunodeficiency virus (HIV). It is biocompatible and well tolerated as it is reabsorbed and degraded in Krebs’s cycle. It can be easily molded, shaped and function as an absorbable space maintainer. This material has lower molecular weight, which permits a more rapid biological degradation, being completely absorbed within 4-6 months.

Fisiograft™ is biocompatible, non-mutagenic, non-allergenic and non-inflammatory and showing osteoconductive properties because it is penetrated and totally substituted by trabecular bone. Animal studies have shown that Fisiograft™ is an osteoconductive degradable copolymer that results information of new bone matrix and trabeculae (10). Post-extraction sockets in mini-pigs has shown that, after 30 days of Fisiograft™ placement, the material was degraded and the sockets were filled with bone, which, in terms of bone volume fraction, trabecular number, and separation, was not statistically different from normal bone (11).

The purpose of this clinical study was to evaluate Fisiograft™ (synthesized co-polymer polylacticpolyglycolic) in the treatment of three wall intrabony defects and to evaluate it clinically as well as radiographically.

Materials and methods

Twenty patients (with 30 defects) diagnosed with generalized chronic periodontitis having two or more three wall vertical defects were selected for this study from the Outpatient Department of Periodontics, Hitkarini Dental College & Hospital, Jabalpur (Madhya Pradesh).

Selection criteria

Inclusion criteria:

1) Patients diagnosed as having generalized chronic periodontitis with probing depth of ≥ 5 mm and radiographic evidence of vertical bone loss;
2) age group of 35-55 years old;
3) patients with good general health, without any history of systemic disease.

Exclusion criteria:

1) Patients showing unacceptable oral hygiene during the presurgical (phase I) period;
2) pregnant women and lactating mothers;
3) smokers;
4) patients with systemically compromised status.

A written informed consent form explaining the nature of the study and surgical procedure was signed by the patients and ethical approval obtained from the local ethical committee from the hospital.

Phase I therapy consisted of Oral Hygiene Instructions, scaling, root planning, and a prescription of chlorhexidine mouth wash. Patients were re-evaluated after phase-1 therapy.
Baseline recording of clinical parameters

Baseline measurements included plaque index, gingival index, probing pocket depth, and clinical attachment level (using a UNC-15 probe with an occlusal stent) (Figure 1).

![Figure 1 - UNC-15 with occlusal stent](image)

Radiographic parameters

An intraoral periapical radiograph of each defect site was exposed using the long cone-paralleling technique using Rinn XCP instrument (Dentsply Rinn, Elgin, IL). The mandibular molar region was the selected site for the study. Exposures were made at 70 KVP, 8 ma, 0.6 sec with inherent filtration of 2 mm AL. Kodak E-speed plus films. The film-to-object and focal spot-to-object distances were each standardized to 20 cm. Digitized images were displayed on the monitor at 5x magnification using Adobe Photoshop 7.0 computer software. A 0.5 mm grid was made on the digitized images and all linear measurements were made using Auto-CAD 2006 software.

Pre-surgical protocol

Following an initial examination and treatment planning discussion, all the selected patients were given detailed instructions regarding the surgical procedure and then subjected to full mouth scaling, root planing and curettage with oral hygiene instructions. Occlusal adjustment was carried out wherever indicated; reevaluation was done after initial therapy.

All the patients were subjected to routine blood examination that included hemoglobin, bleeding time, clotting time, total leucocyte count, differential leucocyte count, and random blood sugar. An ELISA test was also carried out for HIV and hepatitis screening.

All the sites were examined to record the clinical and radiographic parameters.

Surgical protocol

The selected sites were randomly assigned to being either experimental or control sites. After adequate local anesthesia, crevicular incisions were made and the defect site was exposed by reflection of a full-thickness mucoperiosteal flap and debridement of the diseased granulation tissue, followed by thorough root planing and irrigation with normal saline.

At the experimental sites, the defect was filled with Fisiograft™ – Polylactic acid (PLA) and Polyglycolic acid (PGA) particles. The required quantity of graft material was transferred to a dappen dish, mixed with saline, and delivered into the osseous defect incrementally with the help of a Cumine Scaler (Hu-Friedy). The material was placed from the base of the defect coronally to the approximate level of the crest or the remaining osseous walls The operative site was closed with 4-0 black silk sutures and protected with a noneugenol dressing (Figures 2 and 3).

The control sites were left unfilled after surgical debridement, thorough root planning, and irrigation of surgical wound was done with normal saline. The mucoperiosteal flaps were repositioned and secured in place using black, braided (4-0), interrupted silk sutures to obtain primary closure of the interdental space and protected with a noneugenol dressing. All patients were prescribed an analgesic Diclofenac sodium 50 mg, twice a day, and Tetracycline 500 mg thrice a day for five days (Figures 4, 5, 6, and 7).

Post-surgical protocol

After one week following surgery, the dressing and sutures were removed and the surgical site was irrigated thoroughly with saline. As healing was satisfactory and none of the patients experienced any untoward reaction, the recall appointments were made at one month, three, and six months. At each visit, oral hygiene instructions were reinforced and
the surgical sites were professionally irrigated with normal saline.

At the end of three and six months post-therapy, patients were evaluated clinically and radiographically. Clinical parameters (plaque index, gingival index, probing pocket depth, clinical attachment level) and radiographic measurements were repeated for both control and experimental sites (Figures 8 and 9).

Results

The primary goal of periodontal treatment is the maintenance of healthy and comfortable function of natural dentition. When periodontal disease results in a loss of the attachment apparatus, therapy aims at regeneration of the periodontal attachment, which includes the formation of new cementum, functionally oriented periodontal ligament and alveolar bone on the root surface.

The present clinical study was aimed at evaluating the effectiveness of Fisiograft™ as bone grafting material in the treatment of vertical defects in generalized chronic periodontitis patients, and to compare its effectiveness to open flap debridement by itself.

Plaque index

No statistically significant differences were found in the mean values for the plaque index between the test and control groups at baseline (p = 0.170), three months (p = 0.720), and six months (p = 0.180) (Table 1).

![Figure 2 - Flap reflection in control site](image)

![Figure 3 - Suture placed in control site](image)

![Figure 4 - Experimental site measurement with unc-15 & probe](image)

![Figure 5 - Flap reflection and after debridement at experimental site](image)
Gingival index

No statistically significant differences were found in the mean values for the gingival index between the test and control groups at baseline (p = 0.069), three months (p = 0.060), and six months (p = 0.172) (Table 2).

Probing pocket depth

No statistically significant differences were found between the test and control groups at baseline (p = 0.636) and three months (p = 0.109). However, the mean values at six months (p = 0.014) were highly significant. The decrease in probing depth in the
Table 1 - Mean plaque index before and after treatment

<table>
<thead>
<tr>
<th>Plaque index</th>
<th>Control group</th>
<th>Experimental group</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>% Change from baseline</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Baseline</td>
<td>1.465 ± 0.128</td>
<td>—</td>
<td>1.394 ± 0.136</td>
</tr>
<tr>
<td>3 months</td>
<td>0.844 ± 0.196</td>
<td>42.0%</td>
<td>0.810 ± 0.190</td>
</tr>
<tr>
<td>6 months</td>
<td>0.655 ± 0.174</td>
<td>55.0%</td>
<td>0.575 ± 0.144</td>
</tr>
<tr>
<td>p-value</td>
<td>&lt; 0.000**</td>
<td></td>
<td>&lt; 0.000**</td>
</tr>
</tbody>
</table>

Note: ** = highly significant; * = significant.
Source: Research data.

Table 2 - Mean gingival index before and after treatment

<table>
<thead>
<tr>
<th>Gingival index</th>
<th>Control group</th>
<th>Experimental group</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>% Change from baseline</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Baseline</td>
<td>1.09 ± 0.2134</td>
<td>—</td>
<td>0.9407 ± 0.2190</td>
</tr>
<tr>
<td>3 months</td>
<td>0.6973 ± 0.1102</td>
<td>36.0%</td>
<td>0.6173 ± 0.1129</td>
</tr>
<tr>
<td>6 months</td>
<td>0.5220 ± 0.114</td>
<td>52.0%</td>
<td>0.4713 ± 0.0850</td>
</tr>
<tr>
<td>p-value</td>
<td>&lt; 0.000**</td>
<td></td>
<td>&lt; 0.000**</td>
</tr>
</tbody>
</table>

Note: ** = highly significant; * = significant.
Source: Research data.

The experimental site from baseline to six months post operation was 64.26% as compared to the control group, which showed a decrease of 54.52% (Table 3).

Clinical attachment level

The difference between the mean values for the levels of clinical attachment at baseline (p = 0.65) in the test and control groups was not significant. However, the differences in the mean values of clinical attachment levels at three (p = 0.036) and six months (p < 0.001) were statistically significant. This gain in clinical attachment from the baseline to six months postoperatively was 84.82% for the experimental group and 68.83% for the control group (Table 4).

Amount of bone fill in the defects

For control sites, the statistically significant mean difference in defect fill from the baseline was 4.2000 ± 0.9783 mm (p = 0.212) at three months and 3.8000 ± 0.8619 mm (p = 0.014) at six months. For experimental sites, the statistically significant mean difference in defect fill from baseline was 3.6667 ± 1.0293 mm (p = 0.013) at three months and 2.6333 ± 0.8958 mm (p < 0.001) at six months (Table 5).

The differences in the mean values of the amount of defect fill at baseline (p = 0.925) and at three months (p = 0.157) were not significant but the difference was statistically significant at six months (p < 0.001) between the experimental and control groups.

Discussion

Periodontal surgical procedures have focused on the elimination of soft and hard tissue defects (i.e. probing depths and osseous defects) by regenerating new attachment. When periodontal disease results in a loss of the attachment apparatus, therapy aims at regeneration of periodontal attachment that includes formation of new cementum, functionally oriented periodontal ligament, and alveolar bone on the root surface.
Table 3 - Mean probing depth before and after treatment

<table>
<thead>
<tr>
<th>Probing pocket depth</th>
<th>Control group</th>
<th>Experimental group</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>% Change from baseline</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Baseline</td>
<td>8.07 ± 0.88</td>
<td>—</td>
<td>8.20 ± 0.68</td>
</tr>
<tr>
<td>3 months</td>
<td>4.93 ± 0.70</td>
<td>39.0%</td>
<td>4.47 ± 0.83</td>
</tr>
<tr>
<td>6 months</td>
<td>3.67 ± 0.72</td>
<td>55.0%</td>
<td>2.93 ± 0.80</td>
</tr>
<tr>
<td>p-value (baseline-3 months)</td>
<td>0.000**</td>
<td>0.000**</td>
<td></td>
</tr>
<tr>
<td>p-value (baseline-6 months)</td>
<td>0.000**</td>
<td>0.000**</td>
<td></td>
</tr>
</tbody>
</table>

Note: ** = highly significant; * = significant.
Source: Research data.

Table 4 - Mean clinical attachment level before and after treatment

<table>
<thead>
<tr>
<th>Clinical attachment level</th>
<th>Control group</th>
<th>Experimental group</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>% Change from baseline</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Baseline</td>
<td>6.00 ± 0.85</td>
<td>—</td>
<td>6.13 ± 0.74</td>
</tr>
<tr>
<td>3 months</td>
<td>3.00 ± 0.65</td>
<td>50.0%</td>
<td>2.40 ± 0.83</td>
</tr>
<tr>
<td>6 months</td>
<td>1.87 ± 0.64</td>
<td>69.0%</td>
<td>0.93 ± 0.80</td>
</tr>
<tr>
<td>p-value (baseline-3 months)</td>
<td>&lt; 0.000**</td>
<td>&lt; 0.000**</td>
<td></td>
</tr>
<tr>
<td>p-value (baseline-6 months)</td>
<td>&lt; 0.000**</td>
<td>&lt; 0.000**</td>
<td></td>
</tr>
</tbody>
</table>

Note: ** = highly significant; * = significant.
Source: Research data.

Table 5 - Mean radiographic defect fill before and after treatment

<table>
<thead>
<tr>
<th>Depth of the defect</th>
<th>Control group</th>
<th>Experimental group</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean ± SD</td>
<td>% Change from baseline</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td>Baseline</td>
<td>4.6333 ± 0.8756</td>
<td>—</td>
<td>4.6667 ± 1.0465</td>
</tr>
<tr>
<td>3 months</td>
<td>4.2000 ± 0.9783</td>
<td>09.0%</td>
<td>3.6667 ± 1.0293</td>
</tr>
<tr>
<td>6 months</td>
<td>3.8000 ± 0.8619</td>
<td>18.0%</td>
<td>2.6333 ± 0.8958</td>
</tr>
<tr>
<td>p-value (baseline-3 months)</td>
<td>&lt; 0.212</td>
<td>&lt; 0.013*</td>
<td></td>
</tr>
<tr>
<td>p-value (baseline-6 months)</td>
<td>&lt; 0.014*</td>
<td>&lt; 0.000**</td>
<td></td>
</tr>
</tbody>
</table>

Note: ** = highly significant; * = significant.
Source: Research data.

Periodontal regeneration can be achieved by root surface biomodification, bone grafting and guided tissue regeneration techniques. The topical application of chemical agents to modify the root surface is among one of the earliest reputed clinical approaches to prepare root surface for optimal attachment of periodontal tissue and regeneration. Several agents such as citric acid, fibronectin, ethylene dioamine tetra acetic acid (EDTA), and tetracycline have been shown to result in surface biomodification including detoxification, demineralization and collagen fiber exposure (12).

Regenerative therapy is derived from bone or non-osseous materials, and correction of the loss
of alveolar bone occurs by osteoconduction phenomenon. Guided tissue regeneration is based on the principle of guiding the proliferation of various periodontal tissue components during healing following periodontal surgery to achieve regeneration. Placement of a barrier membrane (resorbable or nonresorbable) between the soft tissue flap and root surface favored the repopulation of the wound area adjacent to the root by regenerative cells originating from the periodontal ligament. Biological mediators to enhance cellular repopulation of the periodontal wound are also available, including peptide sequences, protein preparations, and growth factors to regenerate tissues through the principle of mimicking the natural process of tooth formation with the expectation that the regeneration cascade will proceed spontaneously. Such peptides include endogain, platelet-rich plasma preparation etc (12).

The synthesized co-polymer polyactic/polyglycolic has been used in this study and clinical parameters like plaque index, gingival index, probing pocket depth, and clinical attachment level were compared. An attempt was also made to compare the results radiographically. Clinical parameters were recorded at the baseline, three months, and six months after the operation. The present study was a six months’ follow-up study based on the concept that dimensional alterations of the periodontal tissues resulting from active therapy occur within the first six months. The most reliable outcome for assessing periodontal regeneration is human histological investigation, but this is precluded by practical and ethical constraints due to the associated morbidity.

Comparative analysis of plaque index scores of the control and experimental sites at baseline revealed mean scores of 1.465 ± 0.128 and 1.394 ± 0.136, respectively. At the end of six months, the mean value of the control site decreased to 0.655 ± 0.174 whereas the experimental site decreased to 0.575 ± 0.144. This resulted in a ‘t’ value of 1.371 (p < 0.180), indicating a non-significant difference between the two sites. However, the change in plaque index scores from baseline to six months was significant for both the experimental and control sites, which could be attributed to the rigorous oral hygiene maintenance regime, regular follow-up visits, and reinforcement of oral hygiene instructions for the patients throughout the study period. These results are comparable to previous studies reported by Oreamuno et al. (13).

Comparative analysis of gingival index scores of the control and experimental sites at baseline revealed mean scores of 1.0900 ± 0.2134 and 0.9407 ± 0.2190, respectively (p < 0.069), indicating a non-significant difference between the two sites. At the end of six months, the mean value of the control site decreased to 0.5220 ± 0.1114 whereas that of the experimental site decreased to 0.4713 ± 0.0850 (p < 0.172 between the two sites). This improvement in gingival status could be due to the surgery and frequent supportive therapy provided. Similar findings were reported by Yukna et al. (14).

Comparative analysis of the control and experimental sites at baseline revealed probing pocket depth scores of 8.07 ± 0.88 and 8.20 ± 0.68, respectively (p < 0.636), indicating a non-significant difference between the two sites. At three months postoperatively, the values showed a mean of 4.93 ± 0.70 for the control site and 4.47 ± 0.83 for the experimental site, indicating a slightly significant difference (p < 0.109). At the end of six months, the mean value of the control site decreased to 3.67 ± 0.72 whereas that for the experimental site decreased to 2.93 ± 0.80, resulting in a highly significant difference (p < 0.014) between the two sites. This decrease in probing depth in the control group was less than that of the experimental group, which was also statistically significant. This compares favorably with the studies done earlier by Yukna et al., Kreji et al., Nery et al., Stahl and Forum, and Galgut (15-19).

Comparative analysis of the control and experimental sites at baseline revealed clinical attachment level scores of 6.00 ± 0.85 and 6.13 ± 0.74 respectively (p < 0.650). At three months postoperatively, the values showed means of 3.00 ± 0.65 for the control site and 2.40 ± 0.83 for the experimental site, indicating a significant difference (p < 0.036) At the end of six months, the control site’s clinical attachment level had decreased to 1.87 0.64 whereas the value for the experimental site decreased to 0.93 ± 0.80, indicating a statistically highly significant difference (p < 0.001). The comparable gain in the clinical attachment level of control group could be attributed to the formation of the long junctional epithelium instead of increased bone fill and tissue repair as seen in the experimental group. However, the nature of this attachment could not be elicited as it required histological evaluation. This finding is consistent with those of the studies reported by Nery et al., Galgut, Bowen et al., and Reynolds (18-21).
The depth of the defect was the distance from the alveolar crest to the base of the defect. Comparative analysis of the mean percentage change in defect fill for both the sites revealed a about 43.57% bone fill for the experimental site and 17.98% for the control site. Thus, the experimental site had a higher percentage of defect fill than did the control site, the difference being statistically highly significant. These findings are consistent with those of Nery et al., Stahl and Forum, Galgut, and Meffert et al. (16, 19-23).

**Conclusion**

This new combination of synthetic polymer polylactidepolyglycolic proved to be biocompatible and showed improved healing outcomes as compared with open-flap debridement by itself. These outcomes included a reduction of probing depth and a gain in clinical attachment level and amount of bone fill in the defects. There was a significant reduction in the probing pocket depth and a gain in clinical attachment level in both the experimental and control sites. However, the site implanted with the graft material showed a higher reduction in pocket depth and a higher gain in clinical attachment level compared to the control site. Radiographic assessment showed greater defect fill in the experimental site as compared to the control group, indicating the efficacy of the graft material.

Although FisioGraft™ has shown promising results on clinical and radiographic evaluation, additional long-term studies should be undertaken to obtain more clinical evidence for regular use of this material.

**References**


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